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Evaluation of MicroRNAs in Pediatric Epilepsy

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What is already known on this topic?

- The exact pathophysiology of epilepsy is unclear.
- More than 2000 microRNAs have been defined, and more than half of all known miRNAs are expressed in the brain.
- Recent research showed that microRNAs have a role in epilepsy pathogenesis.

What this study adds to this topic?

- The results showed that the expression of microRNA-155 and microRNA-223 was higher in children with epilepsy.
- The expression of the same microRNAs was higher in drugresistant epilepsy patients than in healthy controls.
- The expressions of microRNA-146a, microRNA-155, and microRNA-223 were higher in drug-resistant patients than in drug-responsive children.

ABSTRACT

Objective: The pathophysiology of epilepsy remains unknown. Recent research has shown that microRNA expression changes in epileptic adults. In the present work, we aimed to identify serum microRNA expression in drug-responsive and resistant children with idiopathic generalized epilepsy.

Materials and Methods: The study included 43 (20 male and 23 female) epilepsy patients and 66 (43 male and 23 female) control subjects. The mean ages of the groups were 113.41 ± 61.83 and 105.46 ± 62.31 months, respectively. Twenty-eight epileptic patients were classified as drug resistant. Thirteen of the controls were the siblings of patients with epilepsy. The study only included children with idiopathic generalized epilepsy who had normal brain magnetic resonance imaging. The serum microRNA expressions (microRNA-181a, microRNA-155, microRNA-146, and microRNA-223) were investigated. Expressions of serum microRNA-181a, microRNA-155, microRNA-146, and microRNA-223 were previously investigated in epilepsy patients and children with febrile seizures. Therefore, these microRNAs were chosen. The expressions of serum levels of microRNAs were determined using quantitative real-time polymerase chain reaction.

Results: The results indicated that the expressions of serum microRNA-155 and microRNA-223 were elevated in epileptic children (P < .05). The expression of the same microRNAs was also elevated in individuals with drug-resistant epilepsy compared to healthy controls (P < .05). microRNA-146a, microRNA-155, and microRNA-223 expressions were higher in drug-resistant patients than in drug-responsive children (P < .05). A logistic regression study determined that an increase of microRNA-155 was a risk for epilepsy, while a decrease of microRNA-146a risk for epilepsy.

Conclusion: Few researchers have investigated the function of microRNAs in the development of childhood epilepsy. Our findings revealed that epilepsy patients have abnormal microRNAexpression.

Keywords: Epilepsy, microRNA, drug resistant, children

INTRODUCTION

MicroRNAs (miRNA) are non-coding RNAs that primarily target messenger RNAs (mRNAs).1

They are valuable markers for epilepsy. The first factor is their enrichment in the brain. Although more than 2000 miRNAs have been identified, more than half of these are expressed in the human brain. Second, the presence of brain-enriched microRNAs in a biofluid such as blood would highly suggest a neurologic brain injury disease.²

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Epilepsy is a chronic neurological disorder with incidence rates in children ranging from 0.5 to 8 per 1000 person-years.³⁻⁷ Epilepsy's specific pathophysiology is still a mystery. It might be linked to glial fibroblast expansion, inflammation, pathological circuit re-formation, and neuronal death. MicroRNAs may nuclear factor-kappa B (NF-kB) be involved in the occurrence and development of epilepsy by regulating these pathological processes.⁸⁻¹¹

Studies revealed that miR-146a regulates the expression of NF-kB, interleukin-1 (IL-1), and interferon- α (INF- α) at the post-transcriptional level and affects the inflammatory reaction after an epileptic seizure. Increased miR-146a levels in the epileptic brain may alleviate inflammation.¹²

MicroRNAs-155 also has a role in the regulation of inflammatory pathways in epilepsy. In children with epilepsy, the expression levels of both miR-155 and tumor necrosis factor- α (TNF- α) are increased.

The increased TNF- α levels act as a feedback loop to regulate miR-155 expression. Thus, these 2 molecules interact to mediate the inflammatory process. Glial cells have supportive and protective effects on neurons and are mainly involved in the material metabolism of neurons. Therefore, their dysfunction may be an important cause of epilepsy.

MicroRNA-155 is abundantly expressed in glial cells, and its expression is significantly increased in the brain tissue of epilepsy patients.¹²

Previous studies have found altered expressions of many different microRNAs in epilepsy, such as miR-34, miR-132, miR-146a, miR-199a, miR-375, miR-10b, miR-29a, and miR125a. 12,14-16 Recently, De Beneditis et all compared microRNA expressions of epilepsy patients with controls and showed that miR-142, miR-146a, and miR-223 were significantly upregulated in patients.

Treatment for epilepsy aims to control seizures, improve patients' quality of life, and prevent epilepsy-related morbidity and mortality. Today, anti-epileptic drugs (AED) with various mechanisms of action are used to treat epilepsy. Despite receiving the best medical care possible and the most potent AEDs, about one-third of children with epilepsy still experience seizures. The results of studies showed that microRNA might have a role in developing drug-resistant epilepsy. Tr.23

On the other hand, microRNA-based treatment modalities are being developed. The leading approach to reducing miRNA expression is antisense oligonucleotides antimRs. Experimental animal research reported that antimRs-based treatment reduces seizure frequency.²⁴

Most of the previous research about the role of miRNAs in epilepsy was conducted in adult patients. There are few studies on this topic done with children. ^{25,26} Recently Liu et al²⁷ reported higher relative expression levels of miR-155 in serum exosomes in children with epilepsy than in controls. It was also found that the expression of miR-155 in children with epilepsy was correlated with the course of the disease and the degree of abnormal electroencephalography (EEG).

In the present study, we aimed to evaluate miRNA expression in children with idiopathic generalized drug-resistant epilepsy and compare seizure-free children with 1 AED and healthy controls.

MATERIALS AND METHODS

Subjects

This prospective study was carried out in 3 different cities between October 1, 2018, and October 1, 2021. The ethics committee of Eskişehir Osmangazi University approved the study (IRB No. 2018/49, Date: 27.08.2018), and a university research grant provided the funding. Written informed consent was obtained from the parents of all the participating children. The children's medical background and demographic details were noted. Detailed physical examinations were performed, including neurological examinations.

EPILEPSY GROUP

Inclusion Criteria

(1) Pediatric neurologists at university hospitals diagnosed with epilepsy; (2) ages 1–18 years old; (3) normal brain magnetic resonance imaging (MRI); (4) patients with idiopathic generalized epilepsy; (5) negative findings from the genetic epilepsy panel, which includes tests for the most common genetic variants (SCNA, KCNQ, etc.); and (6) drug-resistant epilepsy defined as failure of adequate trials of 2 tolerated and appropriately chosen and used AED schedules (whether as monotherapies or in combination) to achieve sustained seizure freedom according to criteria's of International League Against Epilepsy (ILAE);²⁸

Exclusion Criteria

- (1) Patients younger than 12 months and older than 18 years old;
- (2) abnormal MRI findings; (3) children with focal epilepsy; and
- (4) positive genetic mutation in epilepsy panel.

Control Group

Sixty-six voluntary healthy age-matched children were included in the control group. Thirteen of them were siblings of patients.

Collection of Blood Samples

Three milliliters of blood was collected at least 1 week after the last seizure, and the serum was immediately separated by centrifugation and stored at -80° C in a freezer.

Quantitative Real-Time Polymerase Chain Reaction

Expressions of serum miR-181a, miR-155, mir-146, and miR-223 were previously investigated in epilepsy patients and children with febrile seizures.^{29,30} Therefore, these miRNAs were chosen.

Pre-designed primers were used for real-time polymerase chain reaction (PCR) to determine the miRNA expression levels of all samples (Table 1).

Total RNA Extraction

Using a TRIzoITM Reagent kit (Thermo, St. Louis, USA), the manufacturer's instructions extracted total RNA from 200 μL of human serum samples. The samples were examined further following the measurement of the nucleic acid content in the total RNA samples using fluorescence spectrophotometry (Colibri Microvolume Spectrometer, Titertek-Berthold,

Table 1. miRNA Primer List			
miRNA	Target Sequences (5'->3')		
hsa-miR-146a UGAGAACUGAAUUCCAUGGGUU			
hsa-miR-223	UGAUCAGUUUGUCAAAUACCCA		
hsa-miR-155	UUAAUGCUAAUCGUGAUAGGGGUU		
hsa-miR-181	AACAUUCAACGUGUCGGUGAGU		

Germany). RNA concentrations ranged from 100 to 400 $ng/\mu L$, and the total RNA purity was verified using A260/A280 ratios (range 1.81–1.97).

Poly(A) Polymerization and Reverse Transcription

The experiment was carried out in a VeritiTM 384-Well Thermal Cycler using the Poly(A) Polymerase, Yeast (ABM, Canada) and OneScript® Plus cDNA Synthesis (ABM, Canada) kit methods (Life Technologies, United States).

Detection of miRNAs by Quantitative Polymerase Chain Reaction

Expression levels of miRNAs for each sample were determined by using ABI 7500 FAST qPCR System (Life Technologies, United States). Real-time PCR was performed in duplicate for each miRNA and included non-template control. Human miRNAs (miR181, miR155, mir146a, and miR223) designs (Diagen, Ankara, Turkey) were used for the study, and endogenous control small ncRNA SNORD47 was chosen as the internal control for normalization. Analysis of real-time PCR data was made possible by calculating the standard curve and CT values. The fold change data's normal distribution was shown by the logarithmic adjustment, which helped normalize the expression levels in cases of over- and under-expression of genes and lessen the effect of extreme values. 31,32

Statistical Analysis

Fold change medians of miRNA expression levels were shown. Due to the small sample size and Shapiro-test Wilk's results that were not normally distributed, non-parametric tests were conducted. The 2 groups' levels of cytokines and miRNA were compared using the Mann-Whitney *U* test. For further evaluation to determine the effects of independent variables on the dependent variable, a model was constructed with a categorical dependent variable (2 groups), and binary logistic regression analysis was performed. Statistical significance was defined as a *P*-value less than .05. Analysis was done using the Statistical Package for Social Sciences (SPSS), version 15.0 software (SPSS Inc.; Chicago, IL, USA).

RESULTS

Sociodemographic and Clinical Features of the Study Group

The study was conducted with 109 children (43 patients and 66 controls). Sixty-three males and 46 females were included. Sixty-five percent of (28/43) idiopathic generalized epilepsy patients were drug resistant. Among healthy controls, 13 children were healthy siblings of epilepsy patients. The study group's mean age was 108.60 ± 61.96 months. The mean ages of epilepsy patients and controls were 113.41 ± 61.83 (4–218) months and 105.46 ± 62.31 (5–216), respectively. Statistical analysis revealed no significant difference between the mean ages of epilepsy patients and controls (P > .05). In addition, no difference was found between the mean age of resistant and drug-responsive patients (Table 2).

microRNA Expressions

The serum miRNA expression analysis revealed an alteration of miR-155 and miR-223 in epilepsy patients compared to the controls (Table 3).

Considering children with drug-resistant epilepsy, the same miRNAs were higher than controls (P < .05).

The results showed that the expression of serum miR-146a, miR-155, and miR-223 in drug-resistant epilepsy patients was higher than in drug-responsive ones (Table 4).

There was no statistically significant difference in the expression of serum miRNAs between drug-resistant epilepsy patients and their healthy siblings (P > .05).

The logistic regression analysis revealed that an alteration of miR-155 was a positive risk factor for epilepsy, whereas an alteration of miR-146a was a negative risk factor for epilepsy (Table 5).

The results of logistic regression showed that also age does not affect miRNA expression (P > .05).

DISCUSSION

Epilepsy is a chronic neurological disease. The exact etiology of epilepsy is still unclear. There is growing evidence of microRNAs have a role in the pathogenesis of epilepsy.

miRNAs can shape neuronal excitability through modulating the expression of specific ion channels, changing the biophysical and firing properties of individual neurons. MicroRNAs can also target ion channels associated with neuroplasticity.

	Epilepsy Patients 113.41 ± 61.83 (4-218) 20/23		Controls	.52 .08
Age (months) (mean \pm SD; minimum–maximum)			105.46 ± 62.31 (5-216) 43/23	
Gender (boys/girls)				
	Drug Responsive	Drug Resistant		
Disease duration median (range) (years)	1.2 (0.8-2.9)	3.4 (1.1-13)	N/A	N/A
AED treatment				
Monotherapy	13	3	N/A	N/A
Polytherapy	2	25		

Table 3. Serum miRNA 146a, 155, 181, and 223 Expressions				
	Epilepsy patients	Controls		
	Fold Change	Fold Change		
	Median 2 ^{-∆∆C†}	Median 2 ^{-∆∆C†}		
miRNAs	(minimum-maximum)	(minimum-maximum)	P	
miR-146a	-0.99 (-4.88 to 2.18)	-0.43 (-5.88 to 10.21)	>.05	
miR-155	13.37 (10.33 to 16.08)	-4.18 (-14.25 to 17.28)	<.05	
miR-181	-0.60 (-3.55 to 15.31)	0.36 (-5.92 to 7.29)	>.05	
miR-223	3.29 (-0.40 to 6.99)	0.01 (-7.92 to 7.75)	<.05	

Table 4. Serum Expression of miRNAs in Drug-Resistant and Drug-Responsive Patients

	Drug-Resistant Patients	Drug–Responsive Patients	
	Fold Change	Fold Change	
	Median 2 ^{-△△C†}	Median 2 ^{-∆∆Ct}	
miRNAs	(minimum-maximum)	(minimum-maximum)	P
miRNA-146a	-0.09 (-4.88 to 2.18)	-1.81 (-2.76 to -0.02)	<.05
miRNA-155	13.72 (11.08 to 16.08)	12.15 (10.33-15.19)	<.05
miRNA-181	-0.78 (-3.55 to 15.31)	2.16 (2.50-10.48)	>.05
miRNA-223	3.61 (-0.40 to 6.99)	2.07 (-0.16 to 5.32)	<.05

Table 5. Logistic Regression Analysis of Expressions of miRNAs

	-	-	•		
		Standard			95% CI (Minimum-
miRNAs	β	Error	P	OR	Maximum)
miR-181	0.030	0.086	0.727	1.031	0.87-1.221
miR-155	0.303	0.125	0.015	1.354	1.061-1.729
miR-146a	-0.635	0.295	0.031	0.530	0.297-0.945
miR-223	0.279	0.263	0.289	1.322	0.789-2.215
miRNA, microRNA; OR, odds ratio.					

Another mechanism by which miRNAs can modulate circuit function is synaptic scaling via interactions with genes associated with synaptic transmission. Apoptosis, inflammation, and pathological circuit re-formation are other mechanisms contribute to the pathophysiology of epilepsy, in which miRNAs might be involved. Brain injury in patients with epilepsy can promote the release of inflammatory factors and induce inflammatory reactions. These inflammatory factors (e.g., IL-1, INF- α , and TNF- α) can destroy the blood-brain barrier and aggravate damage to the nervous system and also excite neurons and promote repeated seizures.

In the present multicenter study, expression levels of miR181a, miR155, miR146a, and miR223 were searched. The expressions of these miRNAs were previously investigated in epilepsy patients and children with febrile seizures. ^{29,30} Therefore, these miRNAs were chosen. The present research showed an alteration of miR-155 and miR-223 in epilepsy patients compared to the controls.

It was reported that miR-155 is connected to the control of inflammatory pathways in epilepsy. In individuals with temporal lobe epilepsy (TLE), it was discovered that miR-155 TNF- α expression levels were both increased. The elevated TNF- α levels control miR-155 expression through a feedback loop. These 2 molecules, therefore, work together to

mediate the inflammatory process.¹³ On the other hand, miR-155 is expressed in glial cells, and its expression is significantly increased in the brain tissue of epilepsy patients.¹²

Research has demonstrated that miR-155 suppresses several genes necessary for microglia activation, phagocytosis, and inflammatory signaling, enabling the microglia to perform beneficial activities in models of motor neuron disease.³⁴

Cai et al³⁵ have shown that injection of antagomirs of miR-155 improves postictal behavior in animal models of pilocarpine-induced status epilepticus.

MicroRNA-223 was another miR that was found to be altered in epilepsy cases in our study. De Benedittis et al¹⁷ discovered that the expression of 3 microRNAs (miR-142, miR-146a, and miR-223) was increased in patients. According to the results of this investigation, patients' miR-223 expression was also changed. Also, in a recent study, it was discovered that miR-223 expression was increased in people with TLE and animal models of epilepsy.³⁶

We found no difference in the expression of miR-146a and miR-223 levels between patients and controls. A review done by Ghafouri-Fard et al³⁷ reported increased expressions of miR-132, miR-146a, miR-181a, and miR-155 in epilepsy.

Increased miR-146a levels in the epileptic brain may suppress inflammation higher levels of pro-inflammatory cytokines, such as IL1, IL6, and TNF- α , as well as higher expression of miR146a, were seen in an animal model of epilepsy. They proposed that miR146a acts as a negative feedback modulator to regulate the synthesis of these inflammatory cytokines.³⁸

Silencing miR-181a produces neuroprotection against hippocampus neuron cell apoptosis post-status epilepticus in children with temporal lobe epilepsy.³⁹

Our results showed the difference in expressions of miR-146a, miR-155, and miR-223 between drug-responsive and drug-resistant cases. In general, approximately 30% of patients with epilepsy suffer from drug-resistant epileptic seizures despite the use of appropriate drugs. In our study, 65% of patients were drug resistant. This higher ratio might be caused by the centers where research was conducted being tertiary hospitals.

A study comprising 77 drug-resistant and 81 drug-responsive epilepsy patients found that miR-194-5p, -301a-3p, -30b-5p, -342-5p, and -4446-3p were significantly deregulated in the drug-resistant group compared to the drug-responsive group and the control group. Among these 5 microRNAs, miR-301a-3p showed the highest diagnostic value for drug-resistant epilepsy and was inversely correlated with seizure severity.³⁰ An adult investigation found that miR-142, miR-146a, and miR-223 were significantly upregulated in patients, and miR-142 and miR-223 expression levels showed well in distinguishing drugsensitive vs. drug-resistant TLE cases.17 Our results showed that miR-146a expression is significantly altered in drug-resistant patients. Leontariti et al⁴¹ also showed elevated serum levels of miR-146a and miR-134 in drug-resistant patients and found that they were a risk factor for developing refractory epilepsy. Drug resistance is significantly influenced by multidrug transporters,

such as the multidrug resistance gene 1 (MDR1), which in astrocytes encodes a P-glycoprotein. Hypoxia-inducible factor-1- α (HIF-1- α). targets and controls MDR1 expression. According to studies, there is a lower concentration of AEDs in the brain as a result of the coordinated overexpression of HIF-1 and MDR1. Through HIF-1, miRNAs have an impact on the pharmacoresistance pathways.²³

Most of the previous investigations about the role of miRNAs in epilepsy were conducted in adult patients.¹⁴⁻¹⁶ There are few studies on this topic done with children that are limited. 8,17,25,26,42 Elnady et al²⁶ looked at the expression of miR-106b and miR-146a in 20 healthy controls and 30 children with epilepsy. Upregulation of miRNAs was observed in patients more than in controls. A systematic review identified only 10 manuscripts performed with 225 children, and 21 different miRNAs were studied previously²⁴. Recently, Liu et al²⁷ searched serum exosome levels of miR-155, and they reported a higher relative expression level in the pediatric epilepsy group as compared to the healthy control group. They also revealed that the expression of miR-155 in children with epilepsy was correlated with the course of the disease and degree of abnormal EEG. In the present research, the relationship between EEG findings and miRNA expressions was not analyzed.

Expressions of miRNAs might have a role in the pathogenesis of not only epilepsy but also epileptic encephalopathy. A study from Turkey conducted with 54 children revealed that the comparison of the epileptic encephalopathy patients' group with healthy controls revealed the upregulation of miR-324-5p and downregulation of miR-146a-5p, miR-138-5p, and miR-187-3p).⁴³

Although patients with epileptic encephalopathy were excluded in the present study, the results of both researches showed an alteration of expression of miR-146a.

According to our best, all control groups in previous research, including adults and children, were composed of healthy nonrelatives. In the current study, 13 of the 66 healthy controls were siblings of epilepsy patients. This fact supports our study even though there was no statistically significant difference between the expression of miRNAs in drug-resistant epilepsy patients and their unaffected siblings. Frye et al44 searched miRNA expression levels in children with autism spectrum disorder (ASD), their typically developed healthy siblings and unrelated controls. They documented 25 miRNAs were upregulated and 43 miRNAs were down-regulated in ASD cases than their typically developed healthy siblings. It must be remembered that it is nearly impossible to interpret a single miRNA's molecular and biophysical functions fully. One miRNA can target many transcripts, and many different miRNAs can target 1 gene. Similarly, miRNAs can target and be targeted by other aspects of epigenetic mechanisms, such as DNA methylation.

MicroRNA expression varies with age.⁴⁵ An association between miR-223 overexpression and age of onset in drug-resistant patients has been reported previously.¹⁷ This point might explain the differences in results of pediatric and adult research about miRNA expression in epilepsy. Genetic polymorphisms might also affect the role of miRNAs in the pathogenesis of epilepsy.⁴⁶ According to a recent study by Boschiero et al⁴⁷ revealed that

rs2910164 polymorphism in the miR-146a gene might be associated with susceptibility to drug-resistant epilepsy due to the decreased MIR146a expression.

Most of the previous research was conducted in patients with abnormal findings such as cortical dysplasia and mesial tempol sclerosis.^{12,25,48}

A recent study done by Nomair et al⁴⁹ included patients with idiopathic generalized and focal epilepsy and documented that both serum miR-146a and miR132 expressions were significantly higher in patients than healthy controls in both groups. However, there was no significant difference in serum miR-146a and miR-132 relative expressions between generalized and focal groups. We found that the expression of serum miR-146a, miR-155, and miR-223 in drug-resistant epilepsy patients was higher than in drug-responsive ones.

They also reported that miR-146 expression was significantly different between drug-responsive and drug-resistant cases among patients with focal epilepsy.

Only children with normal neuroimaging and negative epilepsy panel were included in the study. We thought that this might be a crucial feature of the present study because these inclusion criteria may allow the research group to be more homogenized. We did not compare the results expressions of miRNA according to epileptic syndromes. This might be limitation of present study.

The present study has some limitations. In the present study, the AEDs of patients were not considered. The opinions on the results of the impact of anti-epileptic therapy on a reliable miRNA expression level are not consistent. Rats were given phenobarbital for 2 weeks by Haenisch et al. ¹⁴ but the measured levels of microRNAs did not significantly alter. However, some studies show that AEDs can impact the concentrations of certain miRNAs. Several miRNAs decrease in response to phenobarbital or valproate. ^{50,51} An animal model analyzed the effect of carbamazepine and diazepam before and after therapy and reported none of the 5 miRNAs showed a statistically significant change. ⁵²

In our research, blood samples were collected during the interictal period. A sampling at several time points in patients with epilepsy is still not very common. There are few studies in which miRNA expression measurements were carried out at least twice. End a sample in patients with TLE which were taken during the seizure-free period (on admission and 24 hours after the seizure). Their results showed that the levels of most miRNAs were different between the 2 samples, which indicated their relationship to epilepsy rather than seizure. An investigation searched changes in miRNA expression before and within 30 minutes, 3–6 hours, 20–8 hours, and 3–6 days after seizure in patients with TLE. Analysis showed that the change was transient and associated with seizures. Si

In conclusion, epilepsy is a chronic neurological disease, and its pathophysiology is unclear. Despite the development of new drugs, some patients could not be seizure–free. The results of the present study revealed that miRNAs–146a, –181a, –155, and

223, especially miR-155 and miR-223, might be involved in epilepsy pathogenesis, and expression levels of miRNAs might be useful in predicting drug resistance.

Ethics Committee Approval: The study was approved by the Ethical Committee of Eskişehir Osmangazi University (IRB Number 2018/13, Date:13.08.2018).

Informed Consent: Written informed consent was obtained from the parents of all the participating children.

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Author Contributions: Concept – K.B.C., D.A.; Design – K.B.C., C.Y.; Supervision – K.B.C., C.Y.; Materials – H.G.T., D.C., E.K., C.N.K.; Data collection and processing – H.G.T., D.C., E.K., C.N.K., K.B.C.; Analysis and interpretation – D.A., C.Y., K.B.C.; Literature review – K.B.C., D.A.; Writing – K.B.C.; Critical review – C.Y., K.B.C.

Declaration of Interests: The authors have no conflict of interest to declare.

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